

**2016 Clam Bayou Study Fact Sheet**  
**Florida Department of Environmental Protection**

Because of recent increases in bacteria levels in Clam Bayou, in particular, and in direct response to concerns expressed by the City of Gulfport, the Florida Department of Environmental Protection (DEP) initiated a study of the Bayou this past February. The study, similar to a 2011 DEP evaluation, will assess Clam Bayou water quality, sediments, and selective area plant and animal life (biota). A critical component of the current study, not available in 2011, is the addition of microbial source tracking analyses using chemical and molecular tracers, summarized in more detail below. As DEP gears up to collect more extensive water, sediment, and fish tissue samples, it has shipped sample kits to Gulfport to conduct preliminary water sampling. In addition to source tracking, DEP will analyze samples for the contaminants evaluated in 2011, for comparison purposes. The comparison is to determine whether restoration projects implemented within the Clam Bayou watershed over the past five years have measurably improved water quality and, if not, how future restoration can be better focused.

Microbial Source Tracking

DEP has developed several sophisticated microbial source tracking tools since 2011 to provide a more complete picture of restoration needs. Tracking efforts to date have focused on identifying waters affected by untreated human fecal waste because it represents the greatest potential human health risk. Two chemical tracers, the artificial sweetener sucralose (Splenda®) and the pain reliever acetaminophen (Tylenol®), are used along with the *Bacteroides* HF183 human marker of fecal waste to identify waters containing untreated human wastewater. DEP has recently added the molecular marker, Gull2, to screen for bird fecal sources (*e.g.*, wading birds, gulls, shorebirds, and pigeons) and is developing additional markers to expand its analytical capabilities using sophisticated Quantitative Polymerase Chain Reaction (qPCR) procedures.

Gulfport staff have collected weekly microbial source tracking samples since February 16, along with their routine bacteria sampling at three monitoring stations (Gulfport Marina, Gulfport Beach, and Gulfport Shores) that will be analyzed by DEP. Beginning the week of March 7, microbial source tracking sampling will be expanded to cover the area reflected in Figure 1. Multiple sampling events will occur during dry and wet seasons, which is necessary for reliable, comparable data. Sampling locations may be added during the study to focus on problem areas, if any.

Figure 1. Clam Bayou Microbial Source Tracking Sampling Locations.



### Water, Sediment, and Fish Tissue Sampling

DEP, with help from the Florida Fish and Wildlife Conservation Commission (FWC), will collect other water, sediment, and fish tissue samples for chemical analysis. DEP's laboratory will analyze water samples (dry and wet season) for a broad suite of nutrients, metals, pesticides, base-neutral/acid extractable organic compounds, and polycyclic aromatic hydrocarbons (PAHs). Sediments will be analyzed for percent organic, particle size, phosphorus, metals, pesticides, and base-neutral/acid extractable organic compounds. Finally, the DEP laboratory will analyze the fish tissue samples for pesticides, metals, and PAHs. Sampling locations for this work are still to be finalized. Proposed sediment and fish tissue sampling locations are identified in Figure 2. Water sampling will likely be co-located with some of the microbial source tracking sampling stations.

Water, sediment, and fish tissue analysis results will be compared to:

1. Results obtained during the 2011 Clam Bayou study.
2. Florida water quality criteria (rule 62-302.530 F.A.C.).
3. Florida coastal water sediment quality benchmarks  
[http://www.dep.state.fl.us/waste/quick\\_topics/publications/documents/sediment/volume1.pdf](http://www.dep.state.fl.us/waste/quick_topics/publications/documents/sediment/volume1.pdf).

The results will be used to assess current conditions and, as noted, determine whether restoration projects implemented since 2011 have improved area water quality.

### Infaunal Benthic Macroinvertebrate Community Sampling

DEP's 2011 Clam Bayou report referenced infaunal benthic macroinvertebrate data from a 2009 report by the Environmental Protection Commission of Hillsborough County (EPCHC). Infaunal benthic macroinvertebrate are organisms that live in bottom sediments. Samples were collected in Clam Bayou in 2008 and the results were compared to Clam Bayou samples collected in 2001 and to Tampa Bay Estuary Program reference sites data. In order to compare those data with current conditions, DEP has asked the EPCHC to include infaunal benthic macroinvertebrate sampling locations in Clam Bayou in this year's sampling plan. If this is not possible, DEP will perform its own sampling (Figure 2). Based on seasonal sampling protocols, the EPCHC uses a fall index period to

collect benthic samples and, thus, these results will not be available until 2017. However, other study results will be made available before the benthic sampling results, as noted below.

### Fish Community Sampling

Fish sampling to examine fish community metrics will not be repeated during the 2016 study. Such analyses require extensive, robust sampling efforts in order to overcome the high level of variability in the data. DEP does not believe such sampling is necessary to inform the current study.

### Reporting

DEP will provide an interim report once all of the water, sediment, and fish analyses have been completed and the results interpreted. A final report will be made available once the benthic invertebrate data has been incorporated into the interim report.

Microbial source tracking results will be communicated to Gulfport more frequently if DEP's analyses indicates that untreated human wastewater sources are present. This communication will ensure public health protection.

Figure 2. Green markers indicate the locations of the sediments and macroinvertebrate samples. The red marker notes the location of the fish tissue samples in Clam Bayou.



## Biological Analysis Report

BSSP-2016-03-01-01

Florida Department of Environmental Protection  
Central Laboratory  
2600 Blair Stone Road  
Tallahassee, FL 32399-2400  
DOH Accreditation E31780

Event Description: **Clam Bayou/City of Gulfport MST**  
Request ID: **RQ-2016-02-29-31**  
Customer: **BSSP**  
Project ID: **CLAM\_BAYOU**

Send Reports to:  
FL Dept. of Environmental Protection  
2600 Blair Stone Rd.  
Tallahassee, FL 32399-2400  
Attn: Nia Wellendorf

For additional information please contact  
Loretta Wolfe - Administrator  
J. Marshall Faircloth - Aquatic Toxicology  
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Jacqueline Savage - Microbiology  
Daisys Matthews - Molecular Biology  
Cheryl Swanson - Taxonomy (Invertebrate & Algal)  
Thekkekalathil Chandrasekhar, PhD, QA Officer  
Phone (850) 245-8177

Certified by: Cheryl A. Swanson, Environmental Manager

Date Certified: 10-MAR-2016 16:17



## Case Narrative

Unless otherwise noted, all samples included in this report were received in accordance with protocols referenced in Chapter 62-160, Florida Administrative Code (F.A.C.). Results published in this report pertain only to the samples as submitted to, and received by the laboratory. All times in this report are adjusted to the applicable Eastern Time Zone (EST or EDT).

Results for the following analytical group are included in this report: Microbiology.

Scientific notation may be used in reporting very large or small values. Values reported using scientific notation will take the form of the following example:  $1.3E+03$ , which is equivalent to  $1.3 \times 10^3$  or 1300.

Unless otherwise noted, analytical values for soil and sediment samples are reported on a dry weight basis, and analytical values for waste and tissue samples are reported on a wet weight basis.

Results for TNI accredited tests met requirements established by The NELAC Institute. A double asterisk (\*\*) is used to indicate an analyte/matrix/method for which the laboratory is not TNI accredited or where accreditation for that field of testing is not applicable.

Any significant anomalies or deviations from established protocols are documented in Non-Conformance Reports, which, where appropriate, are included within this analytical report. Additional comments related to specific analytical tests may be included as remarks following the analytical results for each sample. Such comments and remarks are for informational purposes only and are not intended to convey judgement about the usability of the reported data.

A quality control report on the performance of the test method for the submitted samples is included. Uncertainty associated with the analytical results contained in this report can be estimated from the reported quality assurance results and from published quality control acceptance limits for each analytical test. Matrix quality control results (matrix spike recoveries and matrix sample precision) pertain only to the matrix sample tested and do not necessarily reflect test method performance for other samples.

Typical matrix quality control (QC) measurements may include matrix spike recovery, matrix spike duplicate recovery, matrix spike precision and matrix sample precision. Not all matrix QC results may be available or reportable; where they are not an explanation is provided. Typical reasons for unavailable QC results include, but are not limited to, a) insufficient matrix sample to perform some or all QC measurements; b) analyte concentration in the sample replicated was too low for a meaningful measurement of precision and c) analyte concentration in the matrix sample spiked was too high (relative to the amount of analyte spiked) for a meaningful measurement of recovery. Where matrix QC results are unavailable, other method performance metrics (e.g., LCS recovery, LCS precision, surrogate recovery) may be used to assess performance of the method. Comments explaining any missing QC measurements are not intended to convey any adverse conclusions about the quality of the reported data.

Precision is reported as relative percent difference unless otherwise noted.

Quality Control codes as defined below may be used in this report to indicate results that are associated with one or more quality control elements which did not fall within established test method criteria. Such results may be qualified as estimates using a J qualifier as required by 62-160 F.A.C. Explanations are included in the report for any results that were reported as estimates for other reasons.

QC Codes used in this report may include:

- LCS – Recovery for the batch Laboratory Control Sample (LCS) was outside existing control limits;
- MS – Recovery for the batch matrix spike (MS) was outside existing control limits;
- CCV – Recovery for a continuing calibration verification (CCV) standard was outside existing control limits;
- SUR – Recovery of a surrogate (SUR) for associated analytes was outside existing control limits;
- RPD – The precision, measured as relative percent difference (RPD), of batch replicate measurements was outside existing control limits;
- RSD – The precision, measured as relative standard deviation (RSD), of batch replicate measurements was outside existing control limits;
- SMP – Sample - used precision derived from replicate analyses of a sample;

The following data qualifiers are used, where applicable, in this report as specified in 62-160 F.A.C.

- A - Value reported is the mean of two or more determinations.
- B - Results based on colony counts outside the acceptable range.
- I - The reported value is between the laboratory method detection limit and the laboratory practical quantitation limit.
- J - Estimated value and/or the analysis did not meet established quality control criteria.
- K - Actual value is known to be less than value given.
- L - Actual value is known to be greater than value given.
- N - Presumptive evidence of presence of material.
- O - Sampled, but analysis lost or not performed.
- Q - Sample held beyond normal holding time.
- T - Value reported is less than the criterion of detection.
- U - Material was analyzed for but not detected. The reported value is the method detection limit for the sample analyzed.
- V - Analyte was detected in both sample and method blank.
- X - Too few individuals to calculate SCI value.
- Y - The laboratory analysis was from an unpreserved or improperly preserved sample. The data may not be accurate.
- Z - Colonies were too numerous to count (TNTC).

Quality control information from overflow laboratories may not be included in this report. Please refer to the associated report from the overflow laboratory for additional information.

Sample Location: Gulfport Beach

Collection Date/Time: 02/29/2016 12:15

Field ID: MST-ClamBayou3

Matrix: W-SURF-SLT

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1775737	SOP-PCR01	GULL2-qPCR**	1.4E+04	U	TSC/100 mL	P297944	LCS, MS
		HF183-qPCR**	6.4E+03	U	GEU/100 mL	P297944	

Ref. Method and Comment:  
 SOP-PCR01: No amplification of GULL2 marker.  
 SOP-PCR01: No amplification of HF183 marker.

Sample Location: Osgood Point

Collection Date/Time: 02/29/2016 12:35

Field ID: MST-ClamBayou2

Matrix: W-SURF-SLT

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1775738	SOP-PCR01	GULL2-qPCR**	1.4E+04	U	TSC/100 mL	P297944	LCS, MS
		HF183-qPCR**	6.4E+03	U	GEU/100 mL	P297944	

Ref. Method and Comment:  
 SOP-PCR01: No amplification of GULL2 marker.  
 SOP-PCR01: No amplification of HF183 marker.

Sample Location: Gulfpoint Shores

Collection Date/Time: 02/29/2016 13:05

Field ID: MST-ClamBayou1

Matrix: W-SURF-SLT

Sample ID	Ref. Method	Component	Result	Code	Units	Batch ID	QC Codes
1775739	SOP-PCR01	GULL2-qPCR**	2.1E+04	U	TSC/100 mL	P297944	LCS, MS
		HF183-qPCR**	9.5E+03	U	GEU/100 mL	P297944	

Ref. Method and Comment:  
 SOP-PCR01: No amplification of GULL2 marker. Duplicate was analyzed and result is 2.1E+04 U.  
 SOP-PCR01: No amplification of HF183 marker.

### Quality Assurance Report Method Blank Results

Reference Method: SOP-PCR01  
 Batch ID: P297944

Component	Result	Code	Units
GULL2-qPCR	4.2E+04	U	TSC/100 mL
HF183-qPCR	1.9E+04	U	GEU/100 mL

### Quality Assurance Report Laboratory Control Sample Accuracy

Reference Method: SOP-PCR01  
 Batch ID: P297944

Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
GULL2-qPCR	25.8	68.3	F/P	50 - 175
HF183-qPCR	83.8	112	P/P	50 - 175

### Quality Assurance Report Matrix Spike Accuracy

## Quality Assurance Report Matrix Spike Accuracy

Reference Method: SOP-PCR01  
 Batch ID: P297944

Spiked Sample	Component	% Rec.1	% Rec.2	Pass/Fail	Control Limits
1768089	HF183-qPCR	100	80.5	P/P	50 - 175
1771970	HF183-qPCR	70.3		P	50 - 175
1771971	GULL2-qPCR	113		P	50 - 175
1771971	HF183-qPCR	74.4		P	50 - 175
1771972	GULL2-qPCR	83.2		P	50 - 175
1771972	HF183-qPCR	80.0		P	50 - 175
1773360	HF183-qPCR	82.2		P	50 - 175
1773361	HF183-qPCR	84.1		P	50 - 175
1773753	GULL2-qPCR	77.4		P	50 - 175
1773753	HF183-qPCR	78.7		P	50 - 175
1773754	GULL2-qPCR	76.4		P	50 - 175
1773754	HF183-qPCR	70.7		P	50 - 175
1773755	GULL2-qPCR	22.5		F	50 - 175
1773755	HF183-qPCR	86.3		P	50 - 175
1774775	HF183-qPCR	68.4		P	50 - 175
1775737	GULL2-qPCR	52.3		P	50 - 175
1775737	HF183-qPCR	80.6		P	50 - 175
1775738	GULL2-qPCR	51.9		P	50 - 175
1775738	HF183-qPCR	71.8		P	50 - 175
1775739	GULL2-qPCR	35.1	38.0	F/F	50 - 175
1775739	HF183-qPCR	70.5		P	50 - 175

## Quality Assurance Report Precision

Reference Method: SOP-PCR01  
 Batch ID: P297944

Replicated Lab Sample	Component	% RSD/RPD	Sample/Spike/LCS*	Pass/Fail	Control Limits
1768099	HF183-qPCR	21.8	Spike	P	0 - 25
1775739	GULL2-qPCR	7.88	Spike	P	0 - 25

\* Sample, spike and/or laboratory control sample precision (LCS) is reported.  
 Replicate spike precision may be reported when sample results are below quantifiable levels.

## Quality Assurance Report Surrogates

Lab Sample ID: 1775737  
 Field Sample ID: MST-ClamBayou3

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR01	SKETA-qPCR	92.0	P	50 - 300
SOP-PCR01	SKETA-qPCR	109	P	50 - 300

Lab Sample ID: 1775738  
 Field Sample ID: MST-ClamBayou2

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR01	SKETA-qPCR	96.1	P	50 - 300

### Quality Assurance Report Surrogates

Lab Sample ID: 1775738  
 Field Sample ID: MST-ClamBayou2

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR01	SKETA-qPCR	98.1	P	50 - 300

Lab Sample ID: 1775739  
 Field Sample ID: MST-ClamBayou1

Reference Method	Surrogate	% Rec.	Pass/Fail	Control Limits
SOP-PCR01	SKETA-qPCR	118	P	50 - 300
SOP-PCR01	SKETA-qPCR	113	P	50 - 300
SOP-PCR01	SKETA-qPCR	110	P	50 - 300

### Quality Assurance Report Summary

Ref. Method	Analyte	LCS % Recovery		MS % Recovery			Precision	
				LCS	SMP	MS		
SOP-PCR01	GULL2-qPCR	68.3	25.8	22.5	52.3	51.9		7.88
				35.1				
	HF183-qPCR	112	83.8	71.8	70.5	100		21.8
				80.5				

### Reference Method Descriptions

Method / DoH Cert #	Description	Associated Samples
SOP-PCR01 / E31780	Detection and quantification of coastal bird specific <i>Catellibacoccus marimammalium</i> genetic marker with quantitative polymerase chain reaction	1775737, 1775738, 1775739
SOP-PCR01 / E31780	Detection and quantification of human-specific HF183 <i>Bacteroides</i> genetic marker with quantitative polymerase chain reaction	1775737, 1775738, 1775739

### Preparation and Analysis Log

Ref. Method	Received Date	Prep Date/Time	Prepared By	Analysis Date/Time	Analyzed By	Associated Samples
SOP-PCR01	03/01/2016	03/01/2016 14:15	Jonathan M. Woods	03/03/2016	Puja Jasrofia	1775737, 1775738, 1775739

Request Number: RQ-2016-02-29-31  
Clam Bayou/City of Gulfport MST

Florida Department of Environmental Protection  
Central Laboratory Submittal Form

Requester: Daisy Matthews  
Requester: Daisy Matthews  
Project ID: CLAM\_BAYOU  
Collected By: Dwayne Stefnak  
Field Report Prepared By: Dwayne Stefnak  
Sampling Agency: City of Gulfport

Location	Collection (comp begin or grab)	Time	Composite end	Bottle Group(s)
RQ-2016-02-29-31 Location: Gulfport Beach Time: <u>12:15</u>	<input checked="" type="checkbox"/> Comp Date <u>2/29/16</u>		Central Date	<u>A</u>
Field Date: 03-01-16	Matrix (type, e.g., Salt, Fresh, etc)	Time	Diss. Oxygen	
Field ID: <u>MST-ClamBayou3</u>	<u>Salt</u>		<input type="checkbox"/> mg/L Total Res. Chlorine	
STORET #	Temp (C)	pH	<input type="checkbox"/> % sat	
Latitude <u>27° 41' 13.42" N</u>	Salinity (ppt)	Comments	<input type="checkbox"/> ft Sp. Conductance (umho/cm)	
Longitude <u>82° 42' 29.90" W</u>			<input checked="" type="checkbox"/> m	
Location	Collection (comp begin or grab)	Time	Composite end	Bottle Group(s)
RQ-2016-02-29-31 Location: Osgood Point Time: <u>12:35</u>	<input checked="" type="checkbox"/> Comp Date <u>2/29/16</u>		Central Date	<u>A</u>
Field Date: 03-01-16	Matrix (type, e.g., Salt, Fresh, etc)	Time	Diss. Oxygen	
Field ID: <u>MST-ClamBayou2</u>	<u>Salt</u>		<input type="checkbox"/> mg/L Total Res. Chlorine	
STORET #	Temp (C)	pH	<input type="checkbox"/> % sat	
Latitude <u>27° 41' 14.90" N</u>	Salinity (ppt)	Comments	<input type="checkbox"/> ft Sp. Conductance (umho/cm)	
Longitude <u>82° 41' 32.51" W</u>			<input checked="" type="checkbox"/> m	
Location	Collection (comp begin or grab)	Time	Composite end	Bottle Group(s)
RQ-2016-02-29-31 Location: Gulfport Shores Time: <u>13:05</u>	<input checked="" type="checkbox"/> Comp Date <u>2/29/16</u>		Central Date	<u>A</u>
Field Date: 03-01-16	Matrix (type, e.g., Salt, Fresh, etc)	Time	Diss. Oxygen	
Field ID: <u>MST-ClamBayou1</u>	<u>Salt</u>		<input type="checkbox"/> mg/L Total Res. Chlorine	
STORET #	Temp (C)	pH	<input type="checkbox"/> % sat	
Latitude <u>27° 44' 27.45" N</u>	Salinity (ppt)	Comments	<input type="checkbox"/> ft Sp. Conductance (umho/cm)	
Longitude <u>82° 41' 35.08" W</u>			<input checked="" type="checkbox"/> m	
Location	Collection (comp begin or grab)	Time	Composite end	Bottle Group(s)
RQ-2016-02-29-31 Location: Gulfport Shores Time: <u>13:05</u>	<input checked="" type="checkbox"/> Comp Date <u>2/29/16</u>		Central Date	<u>A</u>
Field Date: 03-01-16	Matrix (type, e.g., Salt, Fresh, etc)	Time	Diss. Oxygen	
Field ID: <u>MST-ClamBayou1</u>	<u>Salt</u>		<input type="checkbox"/> mg/L Total Res. Chlorine	
STORET #	Temp (C)	pH	<input type="checkbox"/> % sat	
Latitude <u>27° 44' 27.45" N</u>	Salinity (ppt)	Comments	<input type="checkbox"/> ft Sp. Conductance (umho/cm)	
Longitude <u>82° 41' 35.08" W</u>			<input checked="" type="checkbox"/> m	

Relinquished By:	Date/Time	Shipping Method	Received By:	Date/Time
			<u>MSB</u>	<u>3/1/16 9:27</u>

Group: A	Bottle Type:	QPCR-P-250ML	# of Bottles: 6	Preservative:	ICE	Enter Number of Bottles Sent to Lab
	PCR-GULL2					6
	PCR-HF183					

  

Group: A	Bottle Type:	BG-1L	# of Bottles: 3	Preservative:	ICE	Enter Number of Bottles Sent to Lab
	W-SUC-AA-R					3
	W-UHERB-AA					

Date of Request: 08-FEB-2016  
 Created By: MATTHEWS\_D On: 08-FEB-2016  
 Modified By: WATTS\_J On: 08-FEB-2016  
 Customer: BSSP  
 Project: CLAM\_BAYOU  
 Division: Division of Environmental Assessment and Restoration  
 District:  
 Sampling Event: Clam Bayou/City of Gulfport MST

Send Coolers To:

Phone: 727-893-1090  
 City of Gulfport  
 2401 53rd Street South  
 Gulfport, FL 33707  
 Attn: Don Sopak, Public Works Director

Program Module Number:  
 Priority: 3  
 Event Status: S  
 Criminal Investigation: NO  
 Chemistry Request Reviewed By: WATTS\_J On: 08-FEB-2016  
 Biology Request Reviewed By: SWANSON\_C On: 08-FEB-2016  
 Sampling Kit Required: YES Shp on: 18-FEB-2016  
 Sampling Kit Shipped: YES On: 16-FEB-2016  
 Sampling Kit Packed By: SHAIK\_A  
 Preservatives Needed: NO  
 Date to Receive Samples: 29-FEB-2016  
 Received By:

Send Final Report To:

2600 Blair Stone Rd.  
 Tallahassee, FL 32309-2400  
 Attn: Nia Wellendorf

Report Type: Final Only  
 FTP Data: NO  
 QC Report: YES  
 Date Log: NO  
 Authorization Log: NO

Comments:

Bottle Group A (Water) With 3 Samples:

Bottle Type: QPCR-P-250ML	Number of Bottles: 6	Preserved With: ICE
PCR-GULL2	Template:	(SOP-PCR01) Detection and quantification of coastal bird specific <i>Castelliooccus maritimum</i> genetic marker with quantitative polymerase chain reaction
PCR-HF183	Template: DEFAULT	(SOP-PCR01) Detection and quantification of human-specific HF183 <i>Bacteroides</i> genetic marker with quantitative polymerase chain reaction
Bottle Type: BG-1L	Number of Bottles: 3	Preserved With: ICE
W-SUC-AA-R	Template: DEFAULT	(EPA 8321B) Analysis of sucralose in water matrices by HPLC/MS/MS
W-UHERB-AA	Template: DEFAULT	(DEP SOP: LC-001-2 (based on USGS O-2060-01)) Urea herbicides and other chemicals in water matrices by HPLC/MS/MS

\* - The laboratory is not NELAP certified for this analyte/method, or certification is not applicable.

## Log-in Checklist

NO- 2016-03-29-31

Shipping Method: Fedex

Date/Time of Receipt: 3/1/16/9:27

Cooler ID	Cooler Temperature*	No. Sample Containers in Cooler	Evidence Tape				Tracking # (fill out only if waybill copy is damaged)
			Present?		*Intact?		
			Yes	No	Yes	No	
<u>Blue</u>	<u>-1.0</u>	<u>9</u>		<input checked="" type="checkbox"/>			

\*If the temperature of a cooler is above 6.0 °C (or above 10.0 °C for W-1-4-DIOX), or if Evidence Seal is damaged, identify the containers in the affected cooler(s) on back of this form.

\*\*Note: If "No" is checked in any of the fields below, fill out the back of this form with additional details (Field ID, Test IDs, etc.) and generate an NCR.

\*\*Chain Of Custody/ Field Sheet(s) Included? Yes  No

### CONTAINER CHECK

\*\*Evidence Tape on Bottles? Yes  No  If yes, is it intact? Yes  No

\*\*Caps on tight? Yes  No  \*\*Containers intact? Yes  No

\*\*Sufficient Sample Volume: Yes  No

### CHLORINE CHECK (Blue dot on container)

Check the Box marked "Yes" or "No" for the Presence of Chlorine. If Cl detected, an NCR is required.					
Analysis	Yes	No	Analysis	Yes	No
W-1,4 DIOX			W-PC-AA-SF		
W-AHERB-AA (-AHB-AA-R)			W-PCL-SQ (-R)		
W-BNA (-625, -R)			W-PDQUAT		
W-CARB-AA (-CAB-AA-R)			W-PESTNP-R		
W-ENDO-AA			W-PSCL-TQ		
W-GLYPH			W-PSNP-TQ		
W-NP-AA-SF			W-PST-CL-R		
W-PAH (-R)			W-TR-TQ		
W-PCB-R					

### PRESERVATION CHECK (EXCLUDING SAMPLES SENT TO OVERFLOW LABS, "OV-")

\*\*Acid Preserved Samples: pH ≤ 2.0? Yes  No  NA  Init: MLB

\*\*W-1-4-DIOX (Green dot on container) preserved to pH < 4.0? Yes  No  NA  Init: MLB

Coolers Unpacked/Checked by: MLB Date: 3/1/16 NCRs (Y/N)? N

Event ID: BSSP-2016-03-01-01 NCR #

## Log-in Checklist

### Samples with incorrect pH Preservation or presence of Chlorine

Field ID	Bottle Test ID(s)	pH / Cl	Action Taken

### Samples in Coolers with incorrect Preservation or Damaged Evidence Tape

Cooler ID	Field ID	Bottle Test ID(s)

### Bottles Not Intact

Field ID	Bottle Test ID(s)	Loose Cap		Damaged Cap		Damaged Container		Evidence Tape Not Intact
		OK	BR	OK	BR	OK	BR	

### Samples with Insufficient Volume for Analyses

Field ID	Bottle Test ID(s)	Action Taken

## ENCORE SAMPLES

S-VOC-MS samples in Encores must be frozen within 48 hours of collection.

Were Encore samples received? Yes \_\_\_\_\_ No

Date and Time Placed in Freezer \_\_\_\_\_

Were all samples placed in freezer within 48 hours of collection? Yes \_\_\_\_\_ No \_\_\_\_\_

If no, were samples shipped on dry ice? Yes \_\_\_\_\_ No \_\_\_\_\_

**FOR CLEAN LAB USE ONLY – Methyl Mercury Preservation Check:**

Samples Preserved within 48 hours? Yes \_\_\_\_\_ No \_\_\_\_\_

Date and time samples preserved: \_\_\_\_\_

**Additional Comments:**

00131

01000

02258

FedEx.com 1800.Go.FedEx 1800.463.3339

05899095

**FedEx** Package  
EXPRESS US Airbill

Tracking Number: 8094 7802 5012

1 From  
Date: 2/29/16

Sender's Name: City of Gulfport Phone: 772 893-1065

Company: City of Gulfport

Address: 2401 53rd St. S.

City: Gulfport State: FL ZIP: 33707

2 Your Internal Billing Reference

3 To Recipient's Name: SAMPLE RECEIVING Phone: 850 245-8085

Company: FLORIDA DEP/CHEMISTRY SECTION

Address: 2600 BLAINSTONE RD  
No. 62001 89011 001, South of 201 ZIP code

Address: Tallahassee  
Use this line for the RPO location address or the destination of your shipping address.

City: TALLAHASSEE State: FL ZIP: 32399

0121555852



8094 7802 5012

MARKS

Form 0215

4 Express Package Service \*Special Services

- Next Business Day**
- FedEx First Overnight**
- FedEx Priority Overnight**
- FedEx Standard Overnight**

- Packaging**
- FedEx Envelopes**
- FedEx Paks**
- FedEx Boxes**
- Dimer**

5 Special Handling and Delivery Signature Options

- Signature Required**
- Signature Required - Restricted Address**
- Signature Required - Restricted Address and Signature**
- Signature Required - Restricted Address and Signature and Restricted Address Only**

- Direct Signature**
- Indirect Signature**

- No Signature Required**
- Signature Required - Restricted Address**
- Signature Required - Restricted Address and Signature**
- Signature Required - Restricted Address and Signature and Restricted Address Only**

6 Payment Method

- Bill Me**
- Card**
- Account**
- Third Party**
- Credit Card**
- Cash/Check**

Total Packages: 1



0094 7802 5012